Dysfunction of STING Autophagy Degradation in Senescent Nucleus Pulposus Cell Accelerates Intervertebral Disc Degeneration

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Supplementary Figures and Tables



Supplementary Figure 1 Increasing cytoplasmic dsDNA in degenerative NP cells. (A) Immunofluorescence showed the distribution of TOM20 and dsDNA in the cytoplasm after being treated with IL-1 β , hydrogen peroxide, and hydroxyurea. (Scale bars=50 μ m.)



Supplementary Figure 2 Degenerative changes of the intervertebral disc with age. (A) Hematoxylin-eosin (HE) staining and Safranin O-Fast Green (SOFG) staining of 3,12 and 24 months wild-type (WT) mice. (Scale bars=50 μ m.) (B) Histological score of intervertebral discs in mice at 3,12 and 24 months of WT mice. (Scale bar=50 μ m.) Data are expressed as mean±SD. ****p<0.0001).



Supplementary Figure 3 cGAS knock-out protected mice from instability-induced ID and maintained the height of the intervertebral disc.

(A) HE and SOFG staining showed changes in the IVD of the lumbar vertebra instability model in WT and $cGAS^{-/-}$ mice. (Scale bar=50µm.) (B) Quantitative analysis of the histological score in two groups of vertebral instability models. (C) The calculation method of intervertebral disc height (IDH) and DHI%. (D) X-ray showed

the intervertebral disc height (IDH) of lumbar vertebra instability models. **(E)** Immunofluorescence demonstrated that STING expression increased in the IVDs of WT and cGAS^{-/-} mice with instability model. (Scale bar=20 μ m.) (Scale bar=20 μ m.) (Data are expressed as mean±SD. **p<0.01; ***p<0.001; ****p<0.0001).



Supplementary Figure 4 Elevated expression of STING in the degenerative intervertebral disc and rat puncture model.

(A) The protein with the highest correlation with STING was enriched by the Cytoscape software. (B) Single-cell sequencing showed the changes of cGAS, TBK1, and IRF3 in normal and degenerative human NP cells. (C) The STING expression increased in the IVDs of the rat puncture model with or without injection H151 by using immunofluorescence. (Scale bar= 20μ m.)



Supplementary Figure 5 STING deficiency protects mice from age-induced and instability-induced apoptosis of NP cells in IVDs.

(A) The apoptosis of NP cells in WT and STING^{gt/gt} mice of age-induced and instabilityinduced models was detected by TUNEL tests. (Scale bar= 20μ m.) (B) The STING expression changes were detected by immunofluorescence in age-induced and instability-induced models in WT and STING^{gt/gt} mice. (Scale bar= 20μ m.)



Supplementary Figure 6 Up-regulated STING in the degenerative intervertebral disc promotes apoptosis, activation of inflammatory pathways, and increases catabolism.

(A)TUNEL staining showed that the number of apoptotic NP cells increased after upregulated STING activation. (Scale bar= $50\mu m$.) (B) Western blot examined the changes

of phosphorylation of P65, TBK1, IRF3 in NP control cells and STING overexpression NP cells after stimulated with 2'3'-cGAMP, 2'3'-cGAMP+MRT67307 for 2 hrs. (C) Acian blue staining showed the changes in the extracellular matrix (ECM) of NP cells high-density culture after treated with cGAMP, CMA and over-expressing STING+cGAMP (Scale bar=5mm.) (D) Quantitative analysis of the TUNEL-positive NP cells stimulated with 2'3'-cGAMP and diabzi in Figure S5A. (E) Quantitative analysis of the IOD and IOD/area of Alcian blue staining in supplementary figure S5C. (The cells used were the rat nucleus pulposus cell line. Data are expressed as mean \pm SD, *P<0.05; **p<0.01; ***p<0.001; ****p<0.0001 compared with controls).



Supplementary Figure 7 Cytoplasmic dsDNA was significantly increased in ZMPSTE24-/- mice.

(A) Immunofluorescence demonstrated the distribution of dsDNA in IVDs in WT, STING^{gt/gt}, ZMPSTE24^{-/-} and STING^{gt/gt}×zmpste24^{-/-}mice. (Scale bar=50µm.) (B) The STING expression changes were detected by immunofluorescence in ZMPSTE24^{-/-} and STING^{gt/gt}×zmpste24^{-/-}mice. (Scale bar=20µm.)

Supplementary Table Sequences of Primers for Quantitative Real-time PCR.

Primer sequence			
Rat	ACTB	F	AGTGTGACGTTGACATCCGT
Rat	ACTB	R	CTATGGGTCCAGGCTAAGGC
Rat	B2M	F	AAAAGGCCGATCCGTAGTGC
Rat	B2M	R	TCCGGCACTTAGTGTGCATC
Rat	GUSB	F	AAGCCAATTATCCAGAGCGAGT
Rat	GUSB	R	GGCCACAGTGTGTAGGCTTAG
Rat	mtND1	F	ATAAGCGGCTCCTTCTCCCT
Rat	mtND1	R	GAATGGTCCTGCGGCGTATT
Rat	mtCYTB	F	AGCAACCCTAACACGCTTCT
Rat	mtCYTB	R	ATGGGATTTTGTCTGCGTCG
Rat	mtCOX1	F	AGCAGGGATACCTCGTCGTT
Rat	mtCOX1	R	CAAGGACGGCCGTAAGTGAG