Supplementary Information



Supplementary Figure 1: Restorative Effect of ECM1 on the Regulation of Glycolysis and Angiogenesis in PARPi-Resistant Cells by PS VII

A. qRT-PCR is used to assess the expression of ECM1 mRNA in shRORa and shECM1

cells.B-C. CCK-8 assay (B) and colony formation assay (C) are performed to evaluate the effect of shRORa and shECM1 on the proliferation rate of PARPi-resistant ovarian cancer cells treated with PS VII.D. Wound healing assay is conducted to examine the effect of shRORa and shECM1 on the migratory and invasive abilities of PARPiresistant ovarian cancer cells treated with PS VII.E. The impact of shRORa and shECM1 on tube formation in human umbilical vein endothelial cells (HUVECs) is analyzed using conditioned medium derived from PARPi-resistant ovarian cancer cells with shRORa and shECM1 intervention after PS VII treatment.F. The effect of shRORa and shECM1 on glucose uptake, lactate production, ATP production, and NADPH production in PARPi-resistant ovarian cancer cells treated with PS VII is evaluated.G. ECAR and OCR curves are obtained to evaluate the extracellular acidification rate and oxygen consumption rate, respectively, of PARPi-resistant ovarian cancer cells treated with PS VII after shROR α and shECM1 intervention.H. The effect of shROR α and shECM1H. Western blot experiments are performed to investigate the effects of shRORa and shECM1 on PS VII-treated PARPi-resistant ovarian cancer cells in terms of glycolysis, angiogenesis, and metastasis-related proteins. *p < 0.05, **p < 0.01, ***p< 0.001.





Supplementary Figure 2: Recovery Experiment of PS VII in Reversing PARPi Resistance in Ovarian Cancer Cells through the RORα/ECM1/VEGFR2 Signaling Pathway

A. qRT-PCR is used to detect the expression of VEGFR2 mRNA in shVEGFR2 and

ECM1 OE cells.B-C. CCK-8 assay (B) and colony formation assay (C) are performed to evaluate the effect of shVEGFR2 and ECM1 OE on the proliferation rate of PARPiresistant ovarian cancer cells treated with PS VII.D. Wound healing assay is conducted to examine the effect of shVEGFR2 and ECM1 OE on the migratory and invasive abilities of PARPi-resistant ovarian cancer cells treated with PS VII.E. The impact of shVEGFR2 and ECM1 OE on tube formation in human umbilical vein endothelial cells (HUVECs) is analyzed using conditioned medium derived from PARPi-resistant ovarian cancer cells with shVEGFR2 and ECM1 OE intervention after PS VII treatment.F. ECAR and OCR curves are obtained to evaluate the extracellular acidification rate and oxygen consumption rate, respectively, of PARPi-resistant ovarian cancer cells treated with PS VII after shVEGFR2 and ECM1 OE intervention.G. Western blot experiments are performed to investigate the effects of shVEGFR2 and ECM1 OE on VEGFR2 and FAK signaling pathway-related protein phosphorylation in PS VII-treated PARPi-resistant ovarian cancer cells.H. The effect of PS VII intervention on FAK signaling pathway-related protein phosphorylation, glycolysis, and angiogenesis-related proteins in PARPi-resistant ovarian cancer cells. *p < 0.05, **p < 0.01, ***p < 0.001.

Supplementary Figure 3



Supplementary Figure 3: Supplementary Figure 3: PS VII Combined with PARPi Can Effectively Inhibit Ovarian Cancer Tumor Angiogenesis with Good Safety A. Immunohistochemistry (IHC) is performed to evaluate the protein levels of CD31 in

the tumor tissues of each group (all at 100× and 400× magnification).B. Comparison of

H&E staining of liver and kidney tissues is performed among different groups.

Supplementary Figure 4



Supplementary Figure 4: Association of ECM1, VEGFR2 with PARPi Resistance and Prognosis in Ovarian Cancer

A-D. Based on the IHC staining scores, ovarian cancer patients are divided into high and low expression groups for ECM1 and VEGFR2. H represents high; L represents low. The correlation between ECM1 and VEGFR2 expression levels and PARPi resistance is determined.E. Kaplan-Meier analysis is performed to investigate the relationship between ROR α , ECM1, VEGFR2, and progression-free survival in all ovarian cancer patients and PARPi-resistant ovarian cancer patients.



Supplementary Figure 5: Correlation of RORα/ECM1/VEGFR2 with Prognosis in Ovarian Cancer in TCGA Database

A. Kaplan-Meier analysis is performed to investigate the relationship between RORα/ECM1/VEGFR2 and overall survival in ovarian cancer patients in the TCGA

database.B. Kaplan-Meier analysis is performed to investigate the relationship between RORα/ECM1/VEGFR2 and progression-free survival in ovarian cancer patients in the TCGA database.

Prognostic factors	Patients N (%)	Univariate		Multivariate	
	1	P ^a	HR (95 % CI) ^b	Рь	
All patients	232 (100)				
Age (years)					
< 60 (median)	111 (47.9)				
\geq 60 (median)	121 (52.1)	0.255			
Ascites					
Absence	70 (30.1)				
Present	162 (69.9)	0.165			
BRCA mutation					
BRCA1	86 (37.0)				
BRCA2	146 (63.0)	0.074			
RORa expression status					
Positive	106 (45.7)		1.000		
Negative	126 (54.3)	< 0.001	0.586 (0.437-0.632)	< 0.001	
ECM1 expression status					
Negative	106 (45.7)		1.000		
Positive	126 (54.3)	0.005	1.315 (1.156-1.413)	0.028	
VEGFR2 expression status					
Negative	107 (46.1)		1.000		
Positive	125 (53.9)	0.009	1.415 (1.256-1.592)	0.041	
Chemotherapeutic response					
PARPi sensitive	118 (50.9)		1.000		
PARPi resistant	114 (49.1)	< 0.001	3.840 (2.787-5.289)	< 0.001	

Supplementary Table 1. The association between clinicopathological characteristics and overall survival in ovarian cancer patients

Kanplan-Meier survival analysis and Cox proportional hazards regression analysis

^awithout adjustment

^bwith adjustment for ROR α , ECM1, VEGFR2 expressioin status and chemotherapeutic response

Prognostic factors	RORa-low (%)	RORa-high (%)	P ^a	ECM1-low (%)	ECM1-high (%)	P ^a	VEGFR2-low (%)	VEGFR2-high (%)	P ^a
Age	1	1	0.003	1	1	0.025		1	0.004
≤60 (median)	96 (54.2)	111 (56.1)		108 (52.4)	99 (58.6)		178 (57.6)	29 (43.9)	
> 60 (median)	81 (45.8)	87 (43.9)		98 (47.6)	70 (41.4)		131 (42.4)	37 (56.1)	
Ascites			< 0.001			< 0.001			< 0.001
Absence	16 (9.0)	35 (17.7)		36 (17.5)	15 (8.9)		42 (13.6)	9 (13.6)	
Present	161 (91.0)	163 (82.3)		170 (82.5)	154 (91.1)		267 (86.4)	57 (86.4)	
BRCA mutation			< 0.001			< 0.001			< 0.001
BRCA1	141 (79.7)	185 (93.4)		183 (88.8)	143 (84.6)		267 (86.4)	59 (89.4)	
BRCA2	36 (20.3)	13 (6.6)		23 (11.2)	26 (15.4)		42 (13.6)	7 (10.6)	
Chemotherapeutic response			< 0.001			< 0.001			< 0.001
PARPi sensitive	66 (37.3)	96 (48.5)		98 (47.6)	64 (37.9)		138 (44.7)	24 (36.4)	
PARPi resistant	111 (62.7)	102 (51.5)		108 (52.4)	105 (62.1)		171 (55.3)	42 (63.6)	
Recurrence			< 0.001			0.005			0.635
Absence	48 (27.1)	86 (43.4)		83 (40.3)	51 (30.2)		115 (37.2)	19 (28.8)	
Present	129 (72.9)	112 (56.6)		123 (59.7)	118 (69.8)		194 (62.8)	47 (71.2)	
Survival situation			< 0.001			0.011			0.270
survival	48 (27.1)	86 (43.4)		83 (40.3)	51 (30.2)		115 (37.2)	19 (28.8)	
death	129 (72.9)	112 (56.6)		123 (59.7)	118 (69.8)		194 (62.8)	47 (71.2)	

Supplementary Table 2. The relationship between clinicopathological characteristics and RORa, ECM1, VEGFR2 expression in ovarian cancer patients

Two-sided χ^2 test or Fisher's exact test for distributions between negative and positive expression of ROR α , ECM1, VEGFR2

Supplementary Table 3. List of Primers Sequences	

Primer Name	Primer Sequences (5' - 3')		
~DT. 100 DNA	F: AGGAATTCCCAGTAAGTGCG		
qK1: 185 KNA	R: GCCTCACTAAACCATCCAA		
apt. DOD a	F: CTTGCCGTAGGGATGTCTCG		
ϥ κ1: κΟκα	R: GAAGTTCCGTCAGCCCGTT		
~DT. ECM1	F: AGCACCCCAATGAACAGAAGG		
qK1: ECM1	R: CTGCATTCCAGGACTCAGGTT		
aDT. VECED?	F:TGGTGGTGTGGATGGAGAGT		
qK1: VEGFK2	R:TCTGGAGACTGGAGGGAGTC		
	F: TGCTCTAGAATGAATGAGGGGGGCCCCAGGAGACAGTGAC		
KOKű-OE	R: CCGGAATTCTTACCCATCAATTTGCATTGCTGGCTC		
ECM1 OF	F: TGCTCTAGAATGGGGACCACAGCCAGAGCAGCCTTGGTC		
ECMI-OE	R: CCGGAATTCTCATTCTTCCTTGGGCTCAGAGGTGGAGC		
ECM1 WT promotor	F: TGCTCTAGAGCCCATTCACTAACCCAAAG		
ECWII-W I promoter	R: CCGGAATTCAAAGTCAGCAGCAGACCCATA		
ECM1 MUT promotor	F: TCAGTTTCTCTCGCATGCCTTAAGA		
	R: AACATTCTTAAGGCATGCGAGAGA		