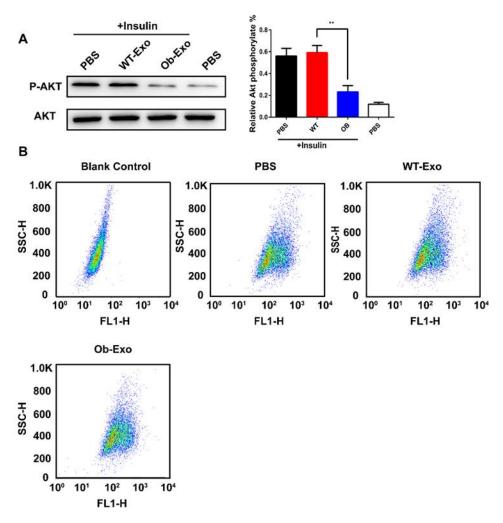
## Ob-exosomes affect the insulin sensitivity and glucose uptake of HepG2 cells

To detect the effect of exosomes on insulin sensitivity and glucose uptake in HepG2 cells, HepG2 cells were treated with WT-exosomes, Ob-exosomes and PBS was used as a negative control. After insulin stimulation, phosphorylation of AKT (Ser473), a central player in the insulin signaling pathway, was detected by Western blot analyses (Supplement Figure 1A). The phosphorylation levels of AKT were lower in the HepG2 cells cultured with Ob-exosomes than in the HepG2 cells cultured with WT-exosomes, suggesting that the insulin responses was inhibited in the HepG2 cells absorbing Ob-exosomes. 2-NBDG, a fluorescent glucose analog was used to detect glucose uptake in HepG2 cells by flow cytometry. The results showed the spontaneous fluorescence intensity of HepG2 cells was 31.10±3.21, the fluorescence intensity of PBS-, WT-exosomes-, Ob-exosomes-treated group was respectively 222.26±1.03, 250.00±2.54, 210.31±3.70. Compared with WT-exosomes-treated group, the fluorescence intensity of Ob-exosomes- treated group was respectively lecreased 15.9% (Supplement Figure 1B). These results showed that insulin-stimulated transport of glucose was inhibited in HepG2 cells treated with Ob- exosomes, indicating that insulin signal transduction in Ob-exosomes-treated HepG2 cells was impaired.



Supplementary Figure 1. Ob-exosomes impair activation of the insulin signaling pathway in vitro. (A) Effect of WT-, Ob-exosomes on AKT phosphorylation of insulin-induced HepG2 cells. TNF- $\alpha$  was used as a positive control and PBS was used as a negative control. \*\*p <0.01, Ob-Exo vs WT-Exo. (B) Flow cytometry dot plots of 10000 HepG2 cells. The 2D plots indicating the relative FL1

fluorescence intensity (2-NBDG) of HepG2 cells. The Blank Control was HepG2 cells without 2-NBDG, showed the spontaneous fluorescence intensity of HepG2 cells. n=3 independent experiments. All values were expressed as means  $\pm$  SD.