

Research Paper

Lnc-MCEI mediated the chemosensitivity of esophageal squamous cell carcinoma via miR-6759-5p to competitively regulate IGF2

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Abstract

Large amounts of long non-coding RNAs (lncRNAs) have been annotated whereas most of them have not been functionally characterized. Here we identified lncRNA ENST00000441932 as an oncogenic lncRNA in esophageal squamous cell carcinoma (ESCC) and named lnc-MCEI (lncRNA mediated the chemosensitivity of ESCC by regulating IGF2). What's more, the effect of lnc-MCEI on the chemosensitivity of ESCC was further evaluated. Bioinformatics analysis demonstrated that lnc-MCEI was involved in the tumorigenesis of ESCC and lnc-MCEI levels were significantly increased in ESCC cells and tissues. Additionally, lnc-MCEI knockdown retarded cell proliferation, colony formation of ESCC cells, but induced cell apoptosis. Moreover, lnc-MCEI knockdown significantly improved the chemosensitivity of ESCC to cisplatin (DDP) both *in vivo* and *in vitro*. Further mechanisms disclosed that lnc-MCEI functioned as a competing endogenous RNA (ceRNA) via sponging miR-6759-5p and IGF2 was a target of miR-6759-5p. Meanwhile, we found that IGF2 suppressed chemosensitivity of ESCC cells via PI3K/AKT pathway. These data suggested that lnc-MCEI was an oncogenic lncRNA and lnc-MCEI knockdown enhanced chemosensitivity of ESCC cells to cisplatin by targeting miR-6759-5p /IGF2/PI3K/AKT axis.

Key words: lnc-MCEI, miR-6759-5p, esophageal squamous cell carcinoma, chemosensitivity, IGF2

Introduction

Esophageal cancer ranks eighth among all tumors and sixth among causes of cancer deaths, with at least 456,000 new cases annually worldwide [1]. The morbidity of esophageal cancer is more than three times higher in men than in women because alcohol abuse and smoking are considered two of the most common risk factors for esophageal cancer and it is more common in less economically developed rural areas [2]. Esophageal carcinoma consists of esophageal squamous cell carcinoma (ESCC) and esophageal adenocarcinoma (EAC) [3]. Compared with EAC, ESCC is the dominant histological type, accounting for up to 80% of all esophageal cancer cases [4]. Cisplatin (DDP) and 5-Fluorouracil (5-FU) have been regarded as the first-line drugs for the

ESCC treatment [5]. Due to the chemoresistance, only 15-25 % of patients survive beyond 5 years despite the combination of surgery with adjuvant therapies such as chemotherapy utilized to treat ESCC [6]. Therefore, it is extremely vital to explore the underlying molecular mechanisms of chemoresistance and develop new strategies to increase the efficacy of chemotherapy drugs.

Long non-coding RNAs (lncRNAs) are considered to be non-coding RNAs with >200 nucleotides (nt) in length and have no protein coding abilities [7]. lncRNAs account for at least 80 % of the human genome and have diversity of biological functions [8]. Emerging studies have demonstrated that lncRNAs are involved in tumorigenesis of

various tumors such as gastric cancer [9], breast cancer [10], osteosarcoma [11], colorectal cancer [11], glioma [12], and so on. Additionally, lncRNAs have been proved to be implicated in the tumorigenesis of esophageal squamous cell carcinoma. Previous studies demonstrated that lncRNA CASC9 promotes ESCC progression with upregulated LACM2 by interaction with CREB-binding protein in the nucleus [13]. Another study indicated that lncRNA CASC9 negatively regulate PDCD4 by binding with enhancer of zeste homolog 2 (EZH2) [14]. In addition, studies showed that lncRNA DUXAP10 promotes tumorigenesis of ESCC by epigenetically silencing p21 [15]. LncRNA GHET1 facilitates the proliferation and invasion of ESCC cells [16]. However, there are still plenty of lncRNAs involved in the ESCC have not been identified. In the present study, the role of lnc-MCEI in the ESCC progression and chemoresistance was evaluated.

Thus, our study was aimed to explore the role of lnc-MCEI in the ESCC progression and chemoresistance. Bioinformatics analysis demonstrated that lnc-MCEI might participate in ESCC chemosensitivity of ESCC to cisplatin and further experiments validates this both *in vivo* and *in vitro*. Therefore, this study revealed that lnc-MCEI promotes ESCC tumorigenesis and chemoresistance and was a potential therapeutic target for ESCC.

Materials and Methods

Tissue sample collection

A cohort of tumor and adjacent normal tissues were collected from 50 patients diagnosed with ESCC in the First Hospital of Jilin University from August, 2013 to August, 2018 under the approval of Institutional Review Board and Human Ethics Committee of the hospital. Patients were performed complete surgical resection (esophagectomy) prior to adjuvant chemo- or radiotherapy and ESCC samples were collected. Adjacent normal tissues were taken from the side of the tumor at least 5 cm away from the edge of the tumor. All tissues were stored at -80 °C until use and all the patients have signed the informed consent form.

Bioinformatics analysis

R package *limma* was used to obtain the differential expressed genes (DEGs) from the microarray data (GSE89102) which was available in the Gene Expression Omnibus (GEO) repository [17]. 3225 DEGs were finally identified (BH-corrected *p* value < 0.01). Probe information has been reported previously [17, 18]. Microarray data (GSE104958) was further used to analyze the expression of long non-coding RNA (lncRNA)-MCEI in tissues with

pathological complete response (pCR) compared to non-pathological complete response (non-pCR).

Cell culture, plasmids construction and transient transfection

Human esophageal epithelial cell line (Het-1A) and ESCC cell lines (TE-1, KYSE-30, EC109 and EC9706) were used in the present study. Het-1A cell line was obtained from the American Type Culture Collection (Manassas, VA, United States) and TE-1 cell line was purchased from Chinese Academy of Medical Sciences (Beijing, China). In addition, KYSE-30 and EC109 cell lines were derived from CoBioer (Nanjing, China) and EC9706 cell line was gotten from Shanghai Huiying (Shanghai, China). KYSE-30 or EC109 cells were transfected with the small interfering RNA (siRNA) targeting lnc-MCEI (si-MCEI), miR-6759-5p mimics, miR-6759-5p inhibitor at 50 % confluence. lnc-MCEI and negative control (NC) hairpin RNA (shRNA) sequences were purchased from GenePharma (Shanghai, China) and cloned into the pBS/U6 vector (Addgene) for lnc-MCEI knockdown. Additionally, the IGF2 expression plasmids construction and transient transfection were performed as the protocol previously described [19].

Quantitative real-time polymerase chain reaction (qPCR)

The mRNA levels of target genes were examined by qPCR as the protocol previously described [20]. For mRNA and miRNA, GAPDH and U6 were used as internal controls. Primer pairs of lnc-MCEI, U6 snRNA and GAPDH were purchased from GenePharma (Shanghai, China) and primers of IGF2 used for amplification in this study are:

- 5'-CGCCGAACCAAAGTGGATTA-3' (forward) and;
- 5'-GGAGAGACAGAGTGAACGTGA-3' (reverse).

MTT, colony formation, migration, invasion assays and flow cytometry

Cell proliferation were analyzed by MTT assays and cells colonies subjected to different treatments were stained and counted 10-14 days after seeding as previously described [21]. Transwell chambers were used to evaluate the migration and invasion capacities of cancer cells and assays were done as previously described [21]. The percentage of apoptotic cells were determined by an Annexin V-FITC/PI Apoptosis Detection Kit (Sigma-Aldrich). All the experimental sets were performed in triplicate.

Dual luciferase reporter assays

Wild-type (WT) or mutant (MUT) 3'-UTR of lnc-MCEI and IGF2 were cloned into the firefly-tagged pGL3 promoter luciferase vector (Promega, Madison, WI, USA) and then were co-transfected into HEK-293 cells with the Renilla control luciferase vector (Promega). Additionally, cells were co-transfected with miR-6759-5p mimics or miR-6759-5p inhibitor for another 48 hours. Luciferase activities were detected with a dual luciferase assay system (Promega) and results were presented as the ratio of firefly/Renilla.

Western blotting

Total protein extraction and western blotting was performed as previously described [22]. Primary IGF2 antibody (ab18954; 1:1000) and GAPDH antibody (ab181602; 1:2000) were purchased from abcam (MA, USA), and p-AKT (BS4009, 1:1000) and t-AKT (BS1810; 1:1000) were obtained from Bioworld Technology (MN, USA).

RNA immunoprecipitation (RIP) assays

For RIP assays, a Magna RIP™ RNA-Binding Protein Immunoprecipitation Kit (Millipore, USA) was utilized according to the manufacturer's protocols. Then the lnc-MCEI mRNA levels were determined by qRT-PCR.

Immunohistochemistry (IHC)

Immunostaining of Ki67 were carried out as previously described [23]. Briefly, sections were incubated with Ki67 antibody (1:200, Sigma-Aldrich, St. Louis, MO, USA) at 4 °C overnight. Horseradish peroxidase (HRP)-conjugated secondary antibodies were further incubated and visualized with 3, 3'-diaminobenzidine (DAB).

Xenograft mouse model

Twenty-four male BALB/c nude mice (six-week-old and six mice in per group) were randomly divided into four groups including NC, sh-lnc-MCEI, DDP and sh-lnc-MCEI+DDP. 3×10^6 EC109 cells of sh-control or MCEI-shRNA in 100 μ L PBS were administered via subcutaneous injection into the right groin of the nude mice. The mice were intraperitoneally injected of DDP (1 mg/kg) from the second week every 3 days. The formula (volume (mm^3) = (length \times width²)/2) was used for the calculation of tumor volume every 3 days. Mice were sacrificed on day 18 post injection and tumors were harvested and weighed [24]. Experimental procedures were approved by the Institutional Animal Use and

Care Committee of the First Hospital of Jilin University.

Statistical analysis

Results (mean \pm SD) were analyzed with a one-way ANOVA or Student's t-test (SPSS 19.0, Chicago, IL, USA). The correlation between miR-6759-5p and lnc-MCEI in ESCC tissues was analyzed by using Spearman correlation analysis. $P < 0.05$ was considered to be statistically significant.

Results

Up-regulated lnc-MCEI was implicated in the promotion of ESCC progression and chemoresistance

To determine candidate genes involved in esophageal squamous cell carcinoma (ESCC) chemoresistance, a bioinformatics analysis was carried out based on the published microarray data (GSE89102) obtained from the Gene Expression Omnibus (GEO). Differential expressed genes (DEGs) were then examined between the 5 paired ESCC tissues and normal tissues by using the microarray information and 1747 significantly up-regulated and 1478 significantly down-regulated DEGs were finally identified ($P < 0.01$; **Figure 1A and 1B**). Further analysis was performed and we found that lnc-MCEI was one of most significantly up-regulated genes in ESCC tissues (**Figure 1C**). Moreover, the data series (GSE104958) was utilized and lnc-MCEI was also significantly increased in ESCC tissues with non-pathological complete response (non-pCR) compared to the tissues with pathological complete response (pCR) after chemotherapy (**Figure 1D**). Additionally, data from The Cancer Genome Atlas (TCGA) RNA-Seq database was analyzed to support this and the lnc-MCEI levels in tumors were significantly higher than normal tissues (**Figure 1E**). Next, the association of lnc-MCEI expression with patient survival was further evaluated according to the TCGA dataset. As shown in **Figure 1F**, increased expression of lnc-MCEI was significantly associated with poor patient survival ($P = 0.17$). Based on the above bioinformatics results, the increased lnc-MCEI expression in ESCC tissues compared to the normal controls or matched normal tissues was confirmed ($N = 50$; **Figure 1G and 1H**). In addition, up-regulated lnc-MCEI expression in ESCC cell lines compared to the human esophageal epithelial cell line (Het-1A) was also observed (**Figure 1I**). These results demonstrated that up-regulated lnc-MCEI was implicated in the promotion of ESCC progression and chemoresistance.

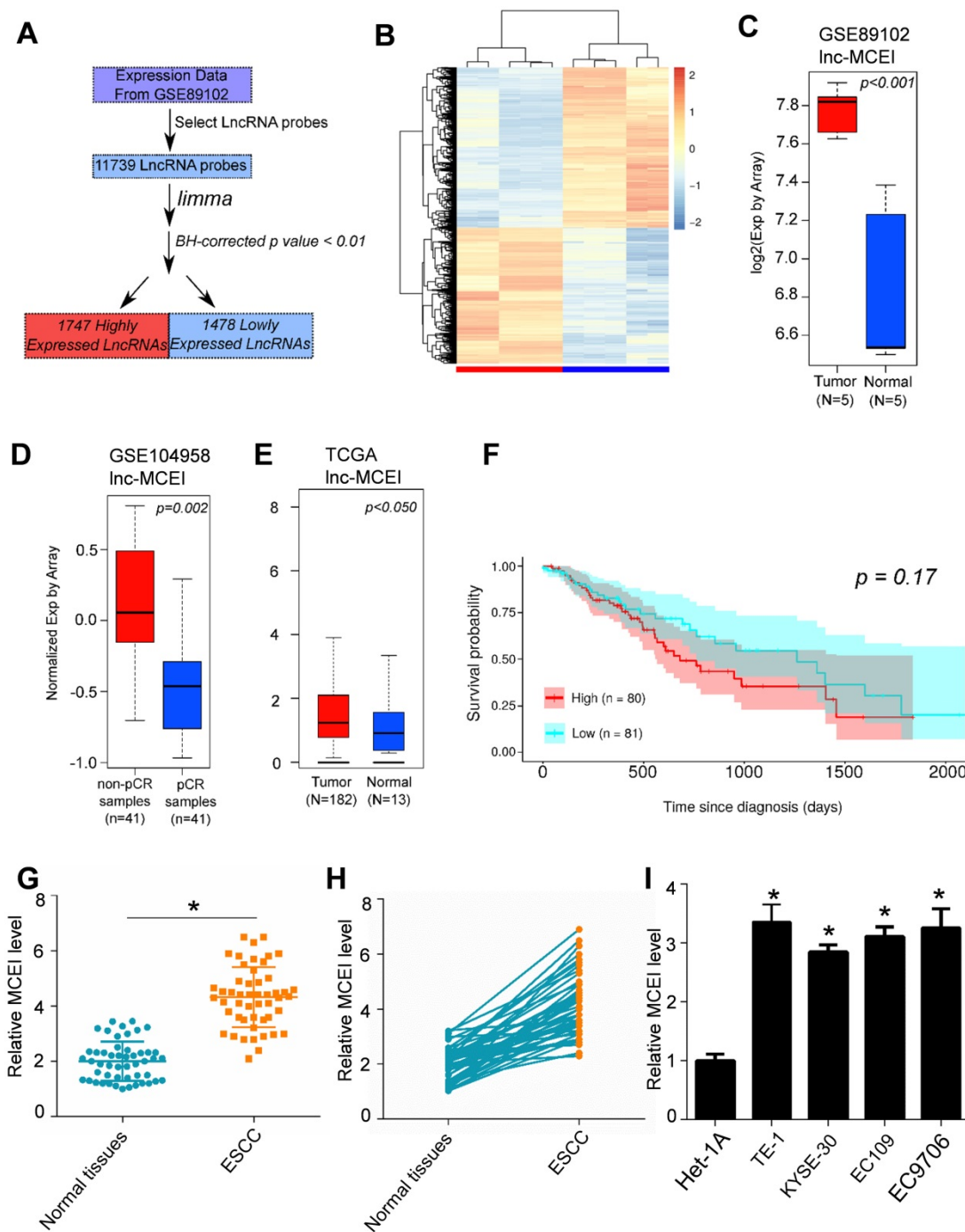


Figure 1. Lnc-MCEI was up-regulated in the ESCC tissues and tissues with non-pathological complete response (non-pCR). **A.** Flow diagram of data analysis based on the published microarray data (GSE89102) with R package *limma*. BH-corrected p value < 0.01 was considered to be statistically significant. **B.** A heat map demonstrated the significantly differential expressed genes (DEGs). The bottom bar denotes the sample types that the red indicates tumor tissues and the blue indicates normal tissues. **C.** Lnc-MCEI was one of most significantly up-regulated genes in ESCC tissues among the DEGs. **D.** Significantly up-regulated lnc-MCEI in tissues with non-pathological complete response (non-pCR) compared to pathological complete response (pCR). **E.** The significantly higher lnc-MCEI expression in tumor tissues compared with the normal tissues according to data from The Cancer Genome Atlas (TCGA) RNA-Seq database. **F.** Increased expression of lnc-MCEI was significantly associated with poor patient survival ($P = 0.17$). **G.** The significantly up-regulated lnc-MCEI in ESCC tissues were observed compared to the normal controls. $N = 50$. **H.** Lnc-MCEI expression was significantly higher in ESCC tissues than the matched normal tissues. $N = 50$. **I.** Lnc-MCEI was significantly up-regulated in ESCC cell lines compared to the human esophageal epithelial cell line (Het-1A). $*P < 0.05$.

Lnc-MCEI knockdown significantly inhibits the malignant biological behaviors of ESCC cells

Subsequently, the role of lnc-MCEI in tumorigenesis was evaluated by knockdown strategies in KYSE-30 and EC109 cells. As shown in

Figure 2A, the lnc-MCEI expression was examined to confirm the knockdown efficiency in both KYSE-30 and EC109 cells. Then the MTT and colony formation assays were used to examine the effect of lnc-MCEI on cell growth. The results suggested that cell proliferation and clone formation of KYSE-30 and

EC109 cells with lnc-MCEI knockdown were significantly suppressed compared to the negative controls (NC; **Figure 2B-D**). Accordingly, cells with lower lnc-MCEI levels exhibited significantly inhibited migration and invasion properties (**Figure 2E and 2F**). Furthermore, lnc-MCEI knockdown significantly increased the percentage of apoptotic cells in both of the cell lines (**Figure 2G**). These data suggested that lnc-MCEI knockdown significantly

inhibited the malignant biological behaviors of ESCC cells.

lnc-MCEI knockdown sensitizes ESCC cells to DDP

Next, to determine the effect of lnc-MCEI on chemosensitivity, KYSE-30 and EC109 cells were treated with cisplatin (DDP) combined with lnc-MCEI knockdown or not. As shown in **Figure 3A and 3B**,

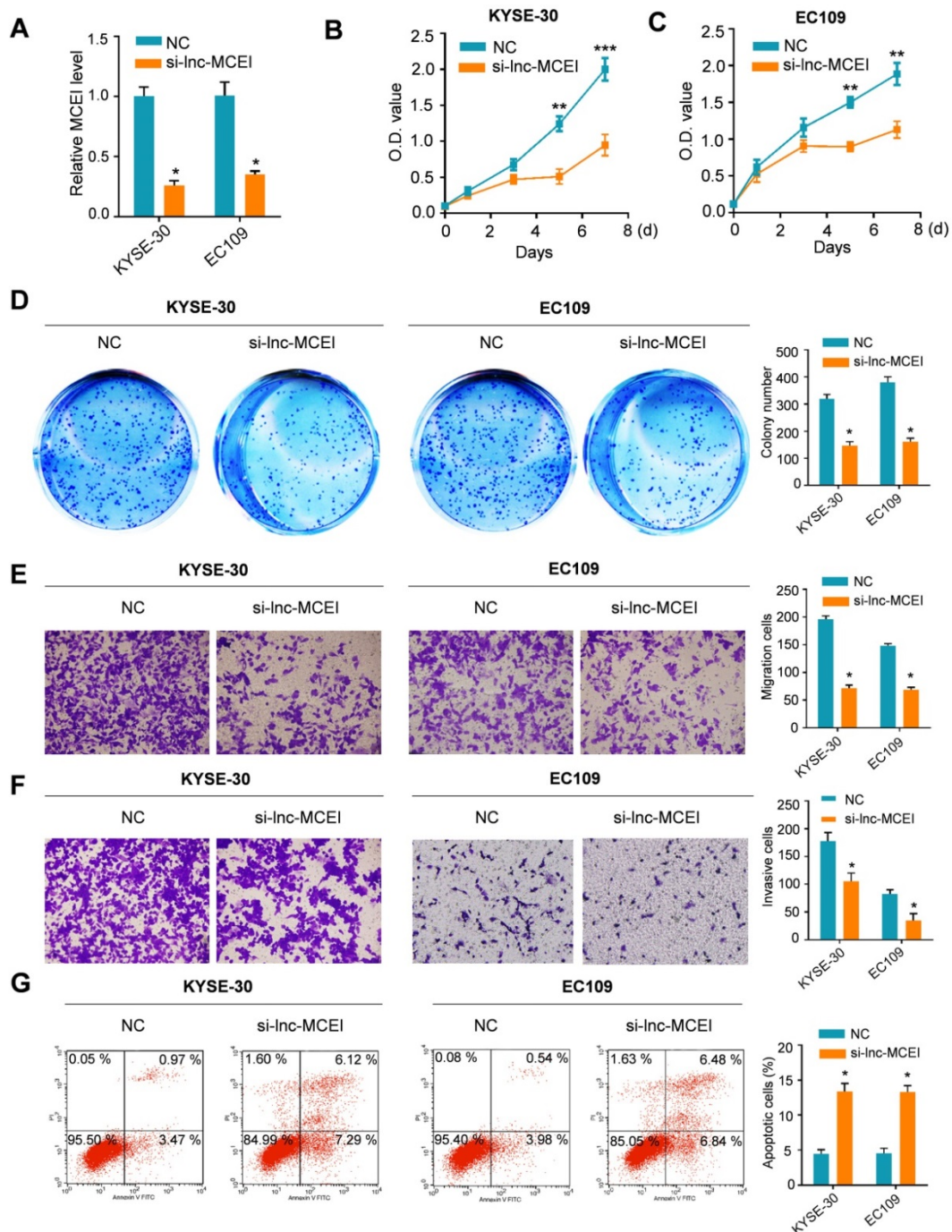


Figure 2. lnc-MCEI knockdown significantly inhibits the proliferation, migration and invasion of ESCC cells. A. lnc-MCEI knockdown was performed in both KYSE-30 and EC109 cells and the knockdown efficiency were detected by qRT-PCR. **B.** MTT assays in KYSE-30 cells. **C.** MTT assays in EC109 cells. **D.** Colony formation assays in both KYSE-30 and EC109 cells. **E.** Migration and invasion assays in both KYSE-30 and EC109 cells. **F.** Invasion assays in both KYSE-30 and EC109 cells. **G.** The results of flow cytometry indicated that lnc-MCEI knockdown significantly increased the percentage of apoptotic cells in both KYSE-30 and EC109 cells. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$.

lnc-MCEI expression significantly increased in a time- and dose-dependent manner in KYSE-30 and EC109 cells subjected to DDP. More importantly, the results of MTT assays and colony formation assays indicated that DDP treatments combined with lnc-MCEI knockdown significantly enhanced the chemosensitivity of KYSE-30 and EC109 cells to DDP (Figure 3C and 3D). In addition, the combination of these two strategies dramatically induced the apoptosis of

KYSE-30 and EC109 cells (Figure 3E). All these data suggested that lnc-MCEI knockdown sensitizes ESCC cells to DDP.

lnc-MCEI functioned as a competing endogenous RNA (ceRNA) via sponging miR-6759-5p

To further explore the underlying mechanisms of lnc-MCEI involved in chemoresistance, the

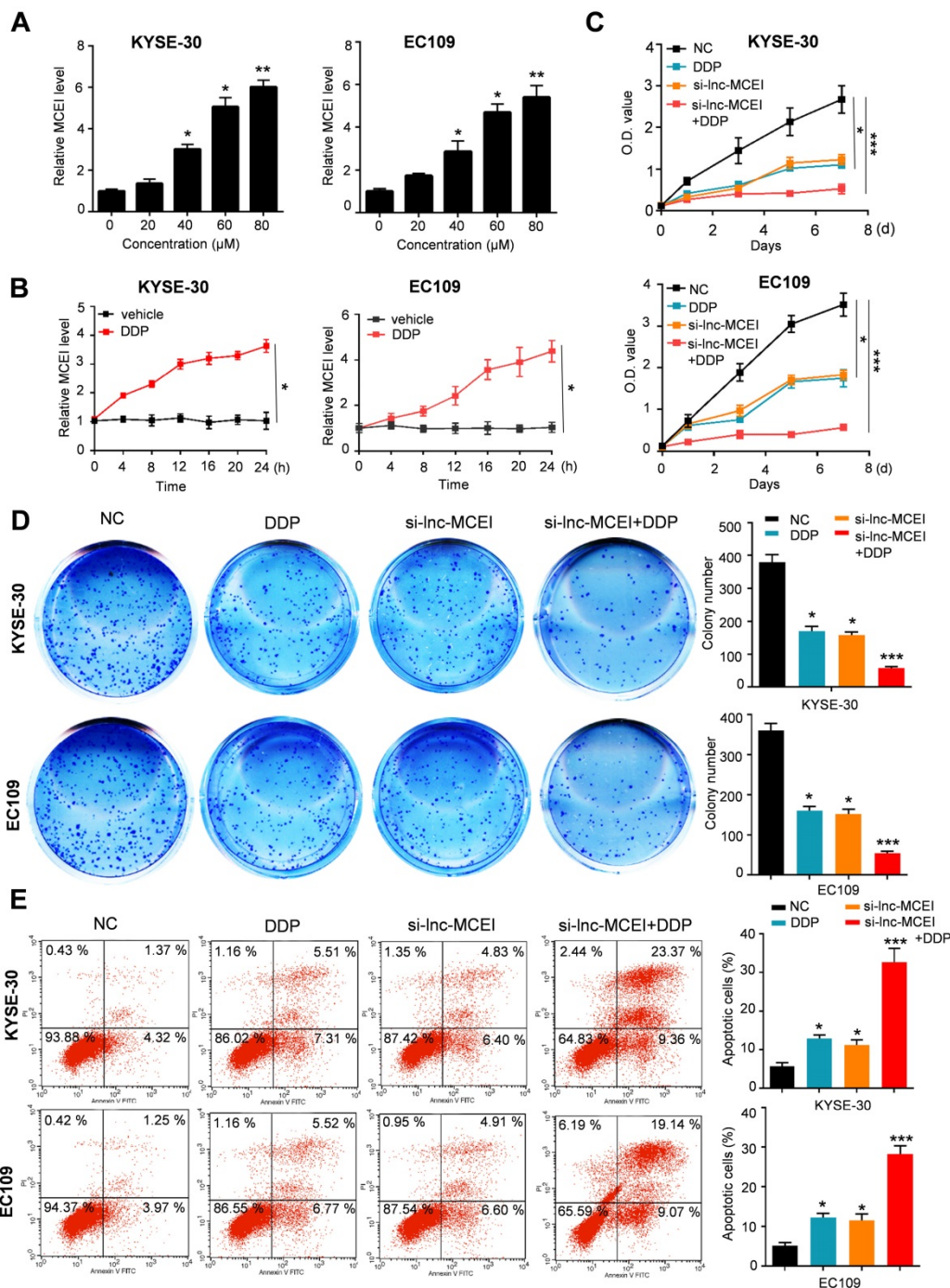


Figure 3. lnc-MCEI knockdown sensitizes ESCC cells to DDP. **A** and **B** lnc-MCEI expression significantly increased in a time- and dose-dependent manner in KYSE-30 and EC109 cells subjected to DDP. **C.** Both KYSE-30 and EC109 cells were treated with si-lnc-MCEI alone or combined with DDP and MTT assays were performed to show the cell proliferation with various treatments. **D.** Colony formation assays in various groups. **E.** The percentage of apoptosis in various groups were tested by flow cytometry. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$.

proposed target genes of lnc-MCEI were examined by searching in LncBase v2.0 database. The results indicated that lnc-MCEI could act as a ceRNA by binding to miR-6759-5p (Figure 4A). In addition, the decreased lnc-MCEI expression in ESCC tissues compared to the normal controls or matched normal tissues was observed (N = 50; Figure 4B and 4C) and lnc-MCEI expression was also decreased in ESCC cell lines compared to the human esophageal epithelial cell line was also observed (Figure 4D). Moreover, we found that there was a significant negative correlation between the expression of lnc-MCEI and that of

miR-6759-5p (Figure 4E). Subsequently, the interaction between lnc-MCEI and miR-6759-5p was confirmed via dual luciferase reporter analysis and the data suggested that miR-6759-5p mimics decreased the luciferase activity of wild-type (WT) lnc-MCEI but had no effect on mutant (Mut) lnc-MCEI (Figure 4F). Additionally, the lnc-MCEI expression was affected by the miR-6759-5p levels in KYSE-30 cells treated with miR-6759-5p mimics or miR-6759-5p inhibitor (Figure 4G). Further RIP assays confirmed that lnc-MCEI functioned as a ceRNA via sponging miR-6759-5p (Figure 4H).

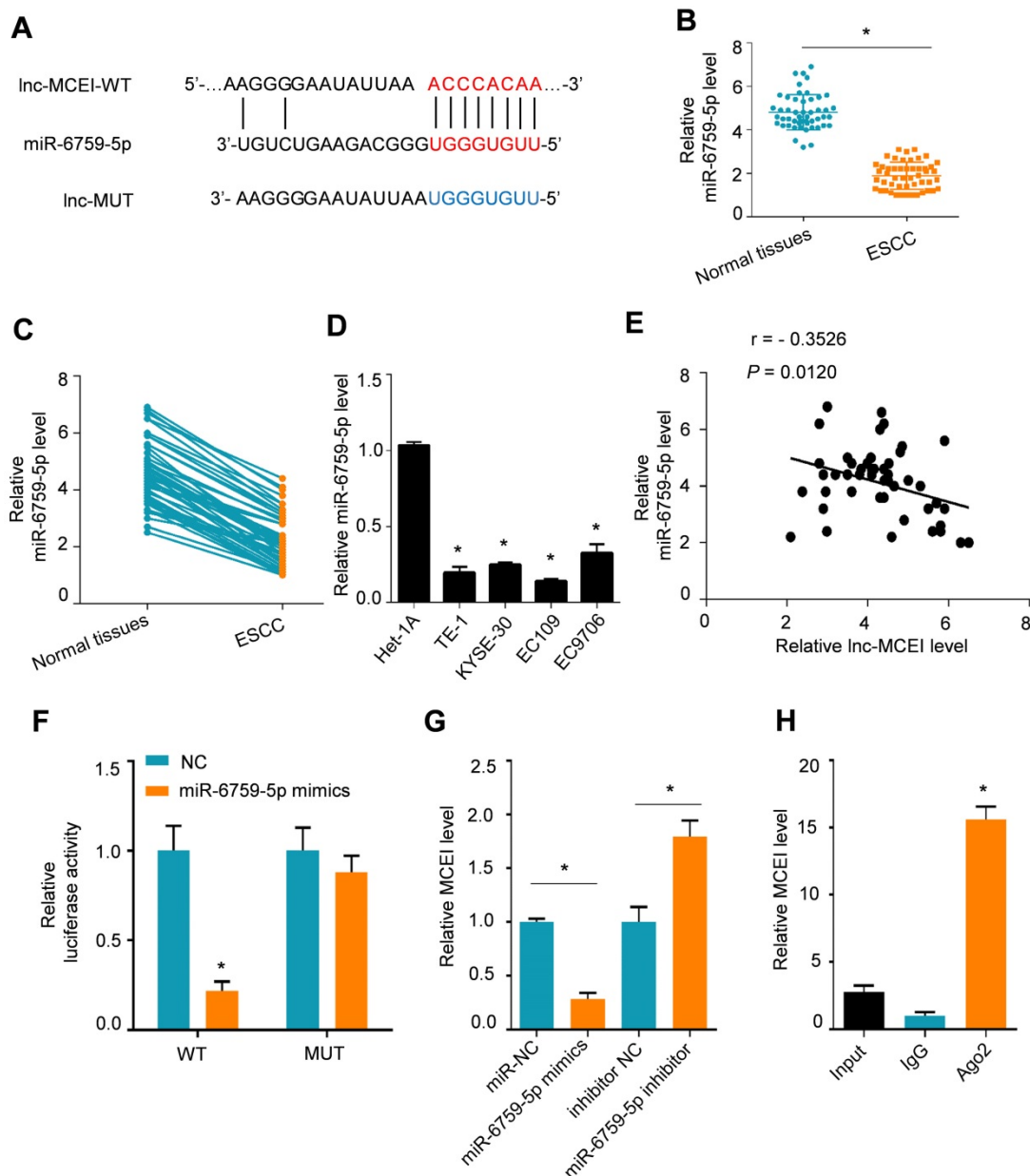


Figure 4. Lnc-MCEI functioned as a competing endogenous RNA (ceRNA) via sponging miR-6759-5p. **A.** The putative binding site of lnc-MCEI and miR-6759-5p was analysed by LncBase v2.0 database. **B.** The significantly down-regulated miR-6759-5p were observed in ESCC tissues compared to the normal controls. N = 50. **C.** The significantly down-regulated miR-6759-5p were also observed in ESCC cell lines compared to the matched normal tissues. N = 50. **D.** miR-6759-5p was down-regulated in ESCC cell lines compared to the human esophageal epithelial cell line (Het-1A). **E.** A significant negative correlation between the expression of lnc-MCEI and that of miR-6759-5p. **F.** Luciferase activity of wild-type or mutated lnc-MCEI in HEK-293T cells examined by the dual luciferase reporter assays. **G.** Relative lnc-MCEI levels in KYSE-30 cells treated with miR-6759-5p mimics or miR-6759-5p inhibitor. **H.** RNA immunoprecipitation (RIP) were performed to examine the interaction between lnc-MCEI and miR-6759-5p. *P < 0.05.

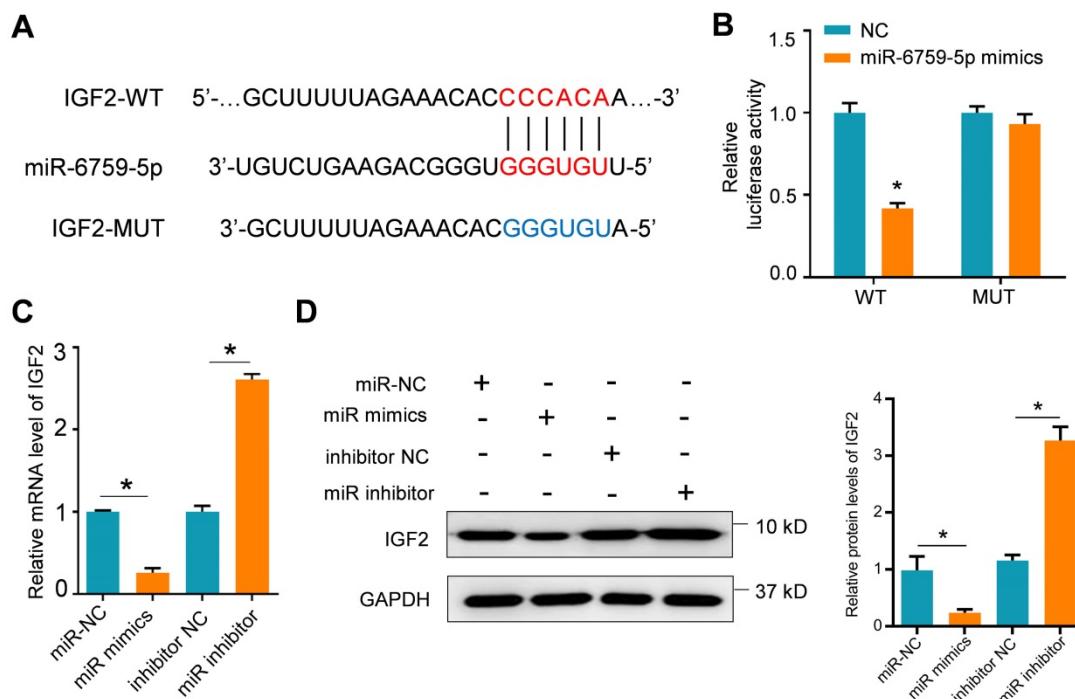


Figure 5. IGF2 is a target of miR-6759-5p. **A.** The putative binding site of lnc-MCEI and miR-6759-5p was analyzed by the TargetScan Human 7.2 database. **B.** Luciferase activity of wild-type or mutated 3'-UTR in HEK-293T cells determined by the dual luciferase reporter assays. **C.** The mRNA expression levels in KYSE-30 cells treated with miR-6759-5p mimics or miR-6759-5p inhibitor. **D.** The protein expression levels of IGF2 in KYSE-30 with the indicated treatments. * $P < 0.05$.

lnc-MCEI mediated the chemosensitivity of ESCC by targeting miR-6759-5p/IGF2/PI3K/AKT axis

The potential target genes of miR-6759-5p were further examined by using the TargetScan Human 7.2 database. The putative binding sites of miR-6759-5p were observed in 3'-UTR of IGF2 mRNA (Figure 5A). The results of dual luciferase reporter assays indicated that the luciferase activity of IGF2 with wild-type (WT) 3'-UTR was significantly decreased whereas IGF2 with mutant 3'-UTR was not influenced after miR-6759-5p mimics treatments (Figure 5B). The mRNA or protein IGF2 expression was also affected by the miR-6759-5p levels in KYSE-30 cells treated with miR-6759-5p mimics or miR-6759-5p inhibitor (Figure 5C and 5D). What's more, lnc-MCEI knockdown significantly decreased the protein expression IGF2 and phosphorylated AKT (p-AKT) but this effect was rescued by miR-6759-5p inhibitor in both KYSE-30 and EC109 cells (Figure 6A and 6B). In addition, MTT assays suggested that IGF2 overexpression rescued the inhibitory effect of the combination of lnc-MCEI knockdown and DDP whereas the effect of IGF2 overexpression was then abolished by the PI3K inhibitor LY294002 in KYSE-30 cells (Figure 6C). Further western blotting data exhibited consistent changes of IGF2 and p-AKT in KYSE-30 cells (Figure 6D). These data indicated that lnc-MCEI mediated the chemosensitivity of ESCC by

targeting miR-6759-5p /IGF2/PI3K/AKT axis.

The miR-6759-5p inhibitor or IGF2 overexpression restores the effect of lnc-MCEI knockdown on chemotherapy

To further confirm the role of miR-6759-5p and IGF2 in the lnc-MCEI-mediated ESCC progression and chemoresistance, colony formation assays were performed and the data indicated that miR-6759-5p inhibitor or IGF2 overexpression rescued the inhibitory effect of the combination of lnc-MCEI knockdown and DDP (Figure 7A). Furthermore, the combination of lnc-MCEI knockdown and DDP significantly induced the apoptosis of KYSE-30 and EC109 cells but the effect was abolished by the miR-6759-5p inhibitor or IGF2 overexpression (Figure 7B). These data suggested that the miR-6759-5p inhibitor or IGF2 overexpression restores the effect of lnc-MCEI knockdown on chemotherapy.

lnc-MCEI inhibition enhances the ESCC chemotherapy to DDP in nude mice

We next examined the role of lnc-MCEI in ESCC tumorigenesis *in vivo*. Immunodeficient nude mice were injected with EC109 cells that had been stably transfected with lnc-MCEI-shRNA (sh-MCEI) or negative control (NC). The results disclosed that tumor volume and tumor weight were significantly decreased in the sh-MCEI and DDP groups and the combination of sh-MCEI and DDP exhibited a more

significant suppressive effect on the tumorigenesis (Figure 8A-C). Moreover, the positive rate of Ki67 was also lower in the sh-MCEI + DDP group than the sh-MCEI or DDP group (Figure 8D). In summary, our data suggested that lnc-MCEI inhibition suppressed ESCC tumorigenesis *in vivo* and lnc-MCEI mediated the chemosensitivity of ESCC by targeting miR-6759-5p /IGF2/PI3K/AKT axis.

Discussion

The mortality of esophageal squamous cell carcinoma (ESCC) remains high despite the rapid promotion in diagnosis and treatment of ESCC [25, 26]. ESCC tumorigenesis has been proved to be regulated by complex mechanisms including cytoplasmic enzymes [27], various oncogenes [28], receptor tyrosine kinases [29], tumor suppressor

genes [30] and lncRNAs [14]. In the present study, we found that lnc-MCEI promotes ESCC tumorigenesis and chemoresistance. lnc-MCEI knockdown significantly inhibits the malignant biological behaviors of ESCC cells and sensitizes ESCC cells to DDP. Further mechanism disclosed that lnc-MCEI functioned as a competing endogenous RNA (ceRNA) via sponging miR-6759-5p and lnc-MCEI mediated the chemosensitivity of ESCC by targeting miR-6759-5p/IGF2/PI3K/AKT axis. Our results suggested that lnc-MCEI was a promising therapeutic target for ESCC.

lnc-MCEI has been rarely studied in previous studies and its role in cancer has never been discovered. Shi-Mei Zhuang et al. reported that lnc-MCEI was one of the candidate lncRNAs frequently amplified in hepatocellular carcinoma

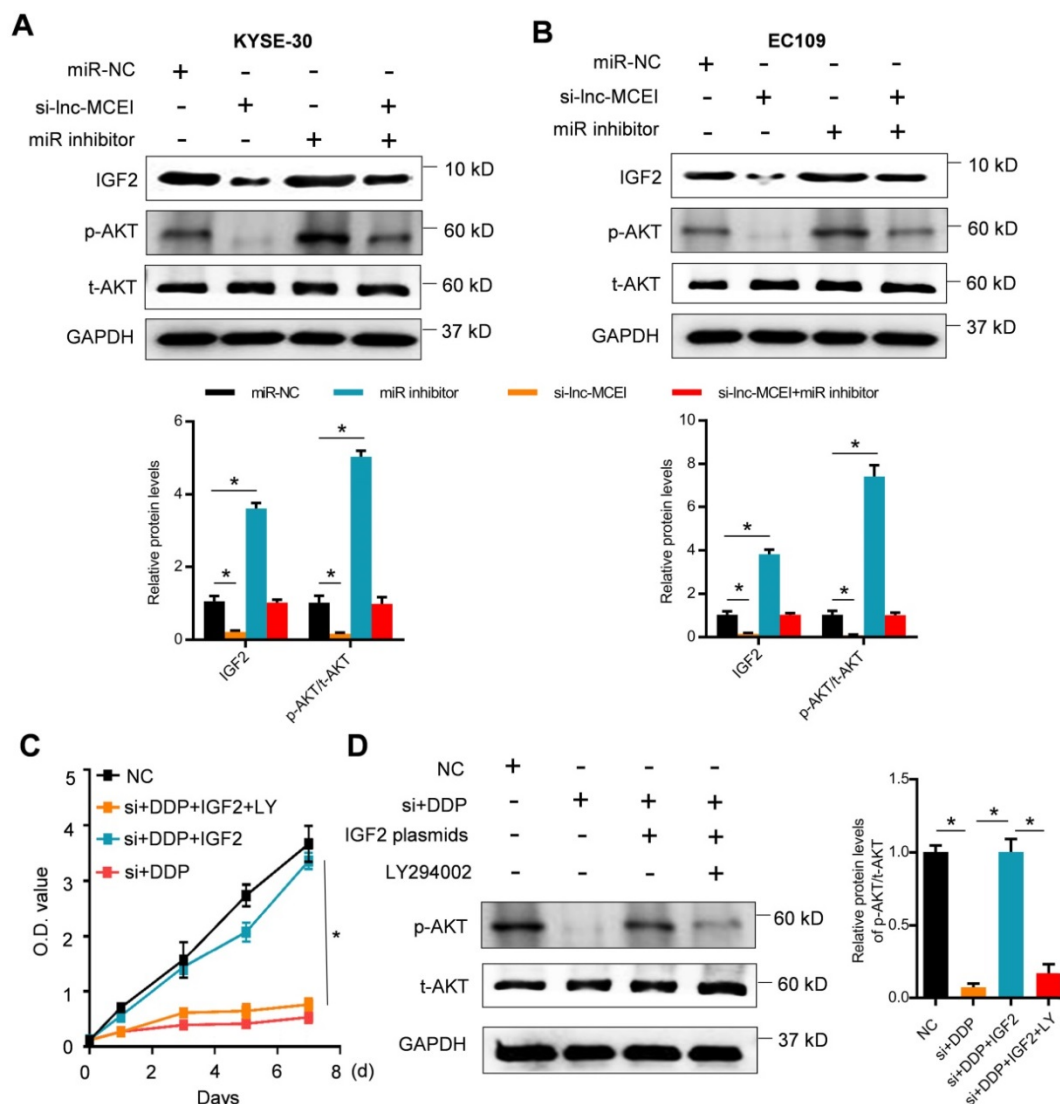


Figure 6. lnc-MCEI mediated the chemosensitivity of ESCC by targeting miR-6759-5p/IGF2/PI3K/AKT axis. **A and B.** The protein expression levels of IGF2, phosphorylated AKT (p-AKT) and total AKT (t-AKT) in KYSE-30 and EC109 cells treated with si-lnc-MCEI alone or combined with miR-6759-5p inhibitor. **C.** MTT assays were performed to examine the cell proliferation in KYSE-30 cells treated with si-lnc-MCEI, DDP, IGF2 plasmids (IGF2) or the PI3K inhibitor LY294002 (LY). **D.** The protein expression levels of p-AKT and t-AKT in KYSE-30 with the indicated treatments. *P < 0.05.

(HCC) but the function in HCC progression have been uncovered [20]. In addition, bioinformatics analysis demonstrated that lnc-MCEI might participate in ESCC tumorigenesis and chemosensitivity based on the published microarray data (GSE89102) which tends to analyze the differential expressed genes (DEGs) between the 5 paired ESCC tissues and normal tissues [17]. Thus, we focused on the role of lnc-MCEI in ESCC and aimed to discover the biological function. Further *in-vivo* and *in-vitro* biological experiments in our study confirm that lnc-MCEI promotes the malignant biological properties and inhibition of lnc-MCEI sensitizes ESCC cells to DDP. Therefore, lnc-MCEI was identified as an

oncogene for the first time in our study.

Studies have shown that lncRNAs may act as a competing endogenous RNA (ceRNA) sponging for miRNAs via the miRNA response elements (MREs) [31-33]. The present results in this study indicated that lnc-MCEI could function as a ceRNA by sponging miR-6759-5p. A global microRNA profiling of metastatic conjunctival melanoma demonstrated that miR-6759-5p was upregulated in metastatic primary conjunctival melanoma [34]. However, miR-6759-5p was proved to be downregulated in ESCC tissues and it is involved in the ESCC tumorigenesis. However, the axis of lnc-MCEI-miR-6759-5p was just one of the mechanisms involved in the ESCC progression. For

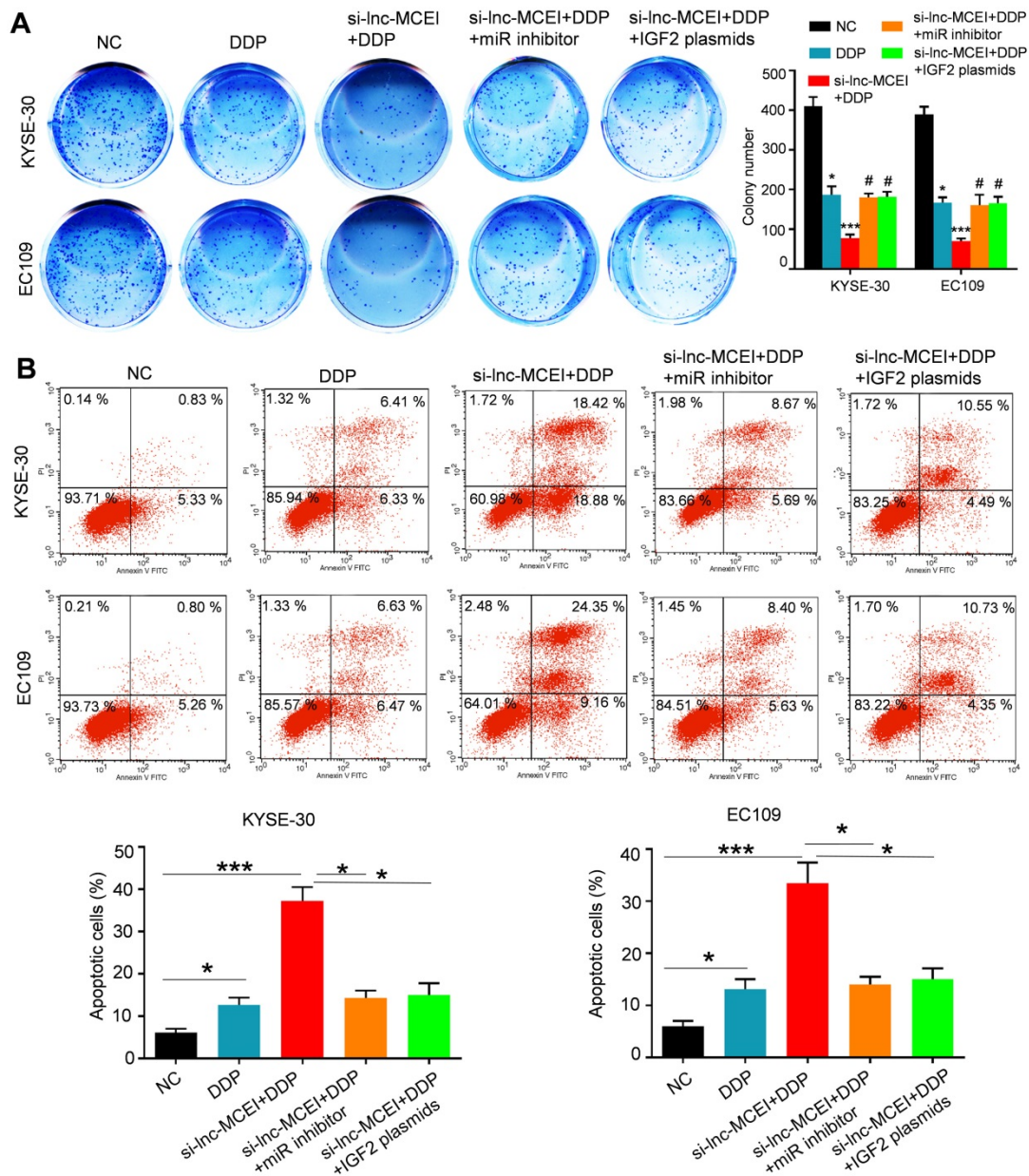


Figure 7. The miR-6759-5p inhibitor or IGF2 overexpression restores the effect of lnc-MCEI knockdown on chemotherapy. A. Colony formation assays in both KYSE-30 and EC109 cells with the indicated treatments (left) and quantification of colony numbers (right). **B.** Flow cytometry was performed to determine the percentage of apoptosis in various groups. **P* < 0.05, ***P* < 0.01 and ****P* < 0.001.

example, bioinformatics analysis also demonstrated that lnc-MCEI may sponge miR-6793-5p. Thus, we will put more effort into the molecular mechanism exploration in our future studies.

IGF2 was increased 1.97-fold in esophageal cancer than that in normal tissues detected by immunohistochemistry (IHC) [35]. Previous studies pointed out that lncRNA 91H had a positive effect on the ESCC by the inhibition of IGF2 [36]. Additionally, IGF2 promotes the stemness of ESCC cells by inducing CD133 expression [37]. Moreover, the epigenetically changes of IGF2 was associated with prognosis of ESCC [38]. Thus, IGF2 promote ESCC tumorigenesis and our data confirm this. In our study,

IGF2 was identified as a target of miR-6759-5p. What's more, studies have revealed that miR-145 sensitizes ESCC to cisplatin via direct inhibition of PI3K/AKT signaling. Therefore, lnc-MCEI/miR-6759-5p/IGF2/PI3K/AKT axis was clarified to be involved in the lnc-MCEI-mediated chemoresistance of ESCC in the present study [24].

Taken together, the beneficial effect of lnc-MCEI knockdown on the malignant biological behaviors of ESCC cells and chemosensitivity of ESCC cells to DDP was verified in the present study. Therefore, lnc-MCEI was identified as an oncogenic lncRNA in ESCC and could be a potential therapeutic target for ESCC by inhibition of miR-6759-5p/IGF2/PI3K/AKT axis.

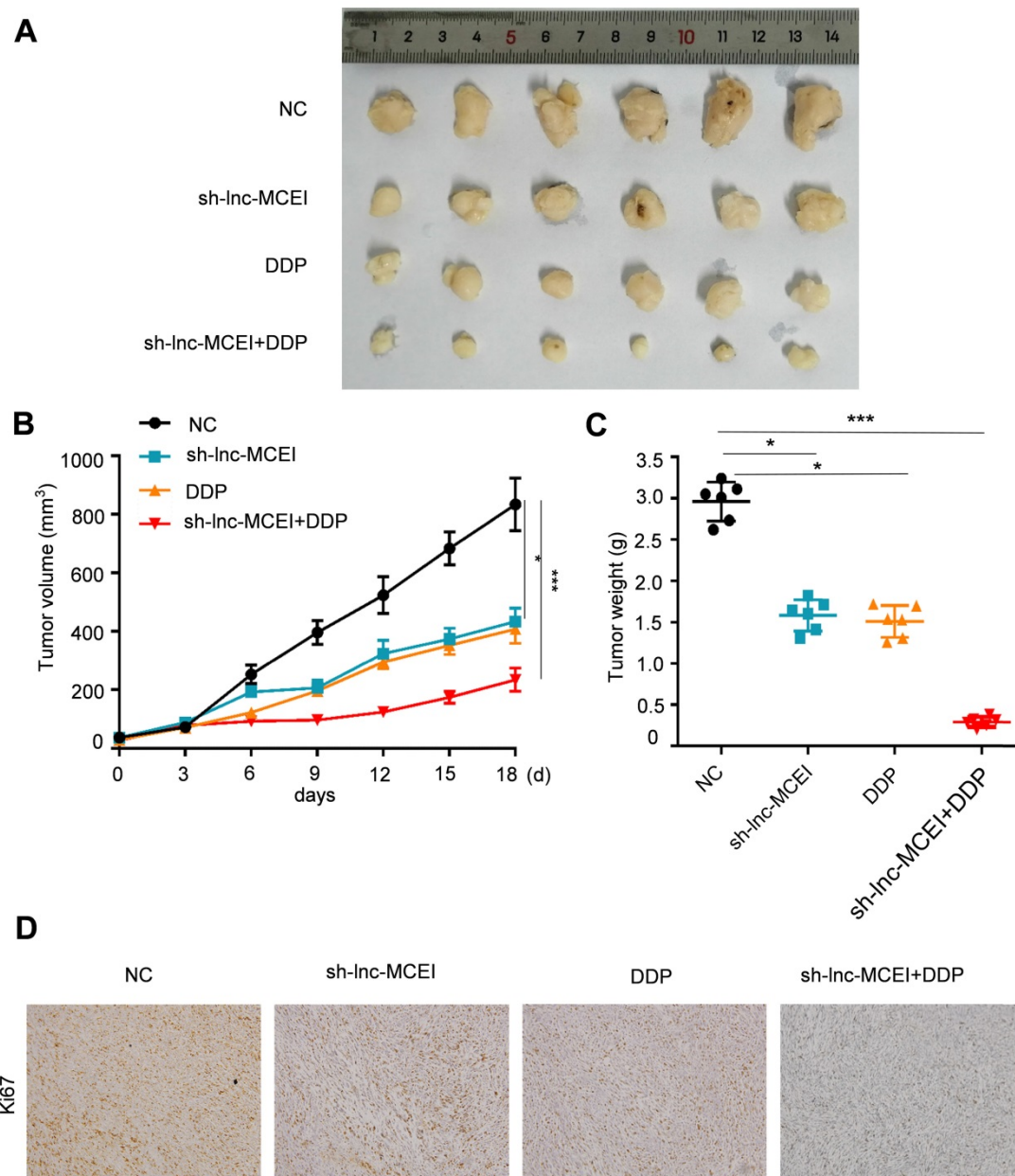


Figure 8. Lnc-MCEI inhibition enhances the ESCC chemotherapy to DDP in nude mice. A. Tumor images dissected from mice treated with sh-lnc-MCEI and DDP alone or combined the both. **B.** Tumor volume in the indicated groups. **C.** Tumor weight in the indicated groups. **D.** Representative images of Ki67 staining in the indicated groups. N = 6; *P < 0.05, **P < 0.01 and ***P < 0.001.

Abbreviations

ceRNA, competing endogenous RNA; DDP, cisplatin; DEGs, differential expressed genes; EAC, esophageal adenocarcinoma; EZH2, enhancer of zeste homolog 2; ESCC, esophageal squamous cell carcinoma; 5-FU, 5-Fluorouracil; GEO, Gene Expression Omnibus; HCC, hepatocellular carcinoma; IGF2, insulin-like growth factor-2; IHC, immunohistochemistry; lncRNAs, long non-coding RNAs; MREs, miRNA response elements; NC, negative control; non-pCR, non-pathological complete response; pCR, pathological complete response; qPCR, quantitative real-time polymerase chain reaction; RIP, RNA immunoprecipitation; shRNA, hairpin RNA; small interfering RNA (siRNA).

Acknowledgements

Authors' contributions

J. P. conceived and designed the study. G. L., W. G. and G. C. performed experimental work. W. L., Y. C. and J. Q. performed data analyses and produced the text and the figures. The final manuscript was approved and contributed by all authors.

Ethics approval and consent to participate

The present study was approved by the Institutional Review Board and Human Ethics Committee of the First Hospital of Jilin University. All the patients have signed the informed consent form prior to their inclusion within the study. All experimental procedures involving animals were conducted in accordance with the Institutional Animal Use and Care Committee of the First Hospital of Jilin University.

Competing Interests

The authors have declared that no competing interest exists.

References

- Zhang Y. Epidemiology of esophageal cancer. *World J Gastroenterol.* 2013; 19: 5598-606.
- Lindkvist B, Johansen D, Stocks T, Concin H, Bjorge T, Almquist M, et al. Metabolic risk factors for esophageal squamous cell carcinoma and adenocarcinoma: a prospective study of 580,000 subjects within the Me-Can project. *BMC Cancer.* 2014; 14: 103.
- Bandla S, Pennathur A, Luketich JD, Beer DG, Lin L, Bass AJ, et al. Comparative genomics of esophageal adenocarcinoma and squamous cell carcinoma. *Ann Thorac Surg.* 2012; 93: 1101-6.
- Domper Arnal MJ, Ferrandez Arenas A, Lanás Arbeloa A. Esophageal cancer: Risk factors, screening and endoscopic treatment in Western and Eastern countries. *World J Gastroenterol.* 2015; 21: 7933-43.
- Ling Y, Chen J, Tao M, Chu X, Zhang X. A pilot study of nimotuzumab combined with cisplatin and 5-FU in patients with advanced esophageal squamous cell carcinoma. *J Thorac Dis.* 2012; 4: 58-62.
- Yang H, Li XD, Zhou Y, Ban X, Zeng TT, Li L, et al. Stemness and chemotherapeutic drug resistance induced by EIF5A2 overexpression in esophageal squamous cell carcinoma. *Oncotarget.* 2015; 6: 26079-89.

- Kung JT, Colognori D, Lee JT. Long noncoding RNAs: past, present, and future. *Genetics.* 2013; 193: 651-69.
- Bartonicek N, Maag JL, Dinger ME. Long noncoding RNAs in cancer: mechanisms of action and technological advancements. *Mol Cancer.* 2016; 15: 43.
- Lin MT, Song HJ, Ding XY. Long non-coding RNAs involved in metastasis of gastric cancer. *World J Gastroenterol.* 2018; 24: 3724-37.
- Zhang T, Hu H, Yan G, Wu T, Liu S, Chen W, et al. Long Non-Coding RNA and Breast Cancer. *Technol Cancer Res Treat.* 2019; 18: 1533033819843889.
- Yang Y, Junjie P, Sanjun C, Ma Y. Long non-coding RNAs in Colorectal Cancer: Progression and Future Directions. *J Cancer.* 2017; 8: 3212-25.
- Rynkeviciene R, Simiene J, Strainiene E, Stankevicius V, Usinskiene J, Miseikyte Kaubriene E, et al. Non-Coding RNAs in Glioma. *Cancers.* 2018; 11: 17.
- Liang Y, Chen X, Wu Y, Li J, Zhang S, Wang K, et al. lncRNA CASC9 promotes esophageal squamous cell carcinoma metastasis through upregulating LAMC2 expression by interacting with the CREB-binding protein. *Cell Death Differ.* 2018; 25: 1980-95.
- Wu Y, Hu L, Liang Y, Li J, Wang K, Chen X, et al. Up-regulation of lncRNA CASC9 promotes esophageal squamous cell carcinoma growth by negatively regulating PDCC4 expression through EZH2. *Mol Cancer.* 2017; 16: 150.
- Wang Z, Ren B, Huang J, Yin R, Jiang F, Zhang Q. lncRNA DUXAP10 modulates cell proliferation in esophageal squamous cell carcinoma through epigenetically silencing p21. *Cancer Biol Ther.* 2018; 19: 998-1005.
- Liu H, Zhen Q, Fan Y. lncRNA GHET1 promotes esophageal squamous cell carcinoma cells proliferation and invasion via induction of EMT. *Int J Biol Markers.* 2017; 32: e403-e8.
- Liang Y, Wu Y, Chen X, Zhang S, Wang K, Guan X, et al. A novel long noncoding RNA linc00460 up-regulated by CBP/P300 promotes carcinogenesis in esophageal squamous cell carcinoma. *Biosci Rep.* 2017; 37.
- Ritchie ME, Phipson B, Wu D, Hu Y, Law CW, Shi W, et al. limma powers differential expression analyses for RNA-sequencing and microarray studies. *Nucleic Acids Res.* 2015; 43: e47.
- Li Y, Fei L, Wang J, Niu Q. Inhibition of miR-217 Protects Against Myocardial Ischemia-Reperfusion Injury Through Inactivating NF-kappaB and MAPK Pathways. *Cardiovasc Eng Technol.* 2020.
- Wang YL, Liu JY, Yang JE, Yu XM, Chen ZL, Chen YJ, et al. lnc-UCID Promotes G1/S Transition and Hepatoma Growth by Preventing DHX9-Mediated CDK6 Down-regulation. *Hepatology.* 2019; 70: 259-75.
- Li Y, Yang Q, Guan H, Shi B, Ji M, Hou P. ZNF677 Suppresses Akt Phosphorylation and Tumorigenesis in Thyroid Cancer. *Cancer Res.* 2018; 78: 5216-28.
- Ruan B, Zhu Z, Yan Z, Yang W, Zhai D, Wang L, et al. Azoramidate, a novel regulator, favors adipogenesis against osteogenesis through inhibiting the GLP-1 receptor-PKA-beta-catenin pathway. *Stem Cell Res Ther.* 2018; 9: 57.
- Simmons ML, Lamborn KR, Takahashi M, Chen P, Israel MA, Berger MS, et al. Analysis of complex relationships between age, p53, epidermal growth factor receptor, and survival in glioblastoma patients. *Cancer Res.* 2001; 61: 1122-8.
- Zheng TL, Li DP, He ZF, Zhao S. miR-145 sensitizes esophageal squamous cell carcinoma to cisplatin through directly inhibiting PI3K/AKT signaling pathway. *Cancer Cell Int.* 2019; 19: 250.
- Chen MF, Chen PT, Lu MS, Lee CP, Chen WC. Survival benefit of surgery to patients with esophageal squamous cell carcinoma. *Sci Rep.* 2017; 7: 46139.
- Zaidi N, Kelly RJ. The management of localized esophageal squamous cell carcinoma: Western approach. *Chin Clin Oncol.* 2017; 6: 46.
- Liu R, Yin L, Pu Y. Association between gene expression of metabolizing enzymes and esophageal squamous cell carcinomas in China. *Genet Test Mol Biomarkers.* 2012; 16: 1211-7.
- Islamian JP, Mohammadi M, Baradaran B. Inhibition of human esophageal squamous cell carcinomas by targeted silencing of tumor enhancer genes: an overview. *Cancer Biol Med.* 2014; 11: 78-85.
- Kashyap MK, Abdel-Rahman O. Expression, regulation and targeting of receptor tyrosine kinases in esophageal squamous cell carcinoma. *Mol Cancer.* 2018; 17: 54.
- Li JS, Ying JM, Wang XW, Wang ZH, Tao Q, Li LL. Promoter methylation of tumor suppressor genes in esophageal squamous cell carcinoma. *Chin J Cancer.* 2013; 32: 3-11.
- Su M, Xiao Y, Ma J, Cao D, Zhou Y, Wang H, et al. Long non-coding RNAs in esophageal cancer: molecular mechanisms, functions, and potential applications. *J Hematol Oncol.* 2018; 11: 118.

32. Wu XS, Wang F, Li HF, Hu YP, Jiang L, Zhang F, et al. LncRNA-PAGBC acts as a microRNA sponge and promotes gallbladder tumorigenesis. *EMBO Rep.* 2017; 18: 1837-53.
33. Cui Y, Yi L, Zhao JZ, Jiang YG. Long Noncoding RNA HOXA11-AS Functions as miRNA Sponge to Promote the Glioma Tumorigenesis Through Targeting miR-140-5p. *DNA Cell Biol.* 2017; 36: 822-8.
34. Mikkelsen LH, Andersen MK, Andreassen S, Larsen AC, Tan Q, Toft PB, et al. Global microRNA profiling of metastatic conjunctival melanoma. *Melanoma Res.* 2019; 29: 465-73.
35. Chava S, Mohan V, Shetty PJ, Manolla ML, Vaidya S, Khan IA, et al. Immunohistochemical evaluation of p53, FHIT, and IGF2 gene expression in esophageal cancer. *Dis Esophagus.* 2012; 25: 81-7.
36. Gao T, He B, Pan Y, Xu Y, Li R, Deng Q, et al. Long non-coding RNA 91H contributes to the occurrence and progression of esophageal squamous cell carcinoma by inhibiting IGF2 expression. *Mol Carcinog.* 2015; 54: 359-67.
37. Xu WW, Li B, Zhao JF, Yang JG, Li JQ, Tsao SW, et al. IGF2 induces CD133 expression in esophageal cancer cells to promote cancer stemness. *Cancer Lett.* 2018; 425: 88-100.
38. Murata A, Baba Y, Watanabe M, Shigaki H, Miyake K, Ishimoto T, et al. IGF2 DMR0 methylation, loss of imprinting, and patient prognosis in esophageal squamous cell carcinoma. *Ann Surg Oncol.* 2014; 21: 1166-74.