Supplementary figures and figure legends



Figure S1. Altered lipids in clinical CRC specimens detected by lipidomics. A, Pie chart of lipid classes whose abundance was significantly changed. B-C, Intensity of phosphatidylcholine (PC) with unsaturated acyl chains (B) and saturated acyl chains (C) from clinical CRC specimens. PM, n=23; nPM, n=22. p < 0.05; **, p < 0.01.



Figure S2. CAFs increased phosphatidylcholine (PC) with acyl chain unsaturation and membrane fluidity in CRC cells. A, Crystal violet staining to quantify the Transwell invasion of the indicated cells after 24 h of exposure to CAF-CM or NF-CM. NF, Normal fibroblast. **B**, Heat map of altered lipids in

CAF-CM-treated HCT116 cells detected by lipidomics. CAF-CM as detected by IncuCyte ZOOM. **C**, ChE (steroid) distribution in CAF-CM-incubated HCT116 cells. n=5. **D**, Intensity of PC containing saturated acyl chains and unsaturated acyl chains in CAF-CM-incubated HCT116 cells. n=5. **E**, Normalized fluorescence recovery curve of HCT116 cells cultured with control/conditioned medium. n = 3. **F**, Fluorescence polarization (Relative Polarization_{corr}) in CRC cells incubated with 25% CAF-CM or a double dose of CAF-CM (50%). n = 2 independent experiments with 2 CAF-CM samples from different CAF cell lines. Bars, mean ± SD. *, p < 0.05, ***, p < 0.001.



Figure S3. C16:0 inhibits CRC cell migration and invasion by decreasing membrane fluidity. **A**, Fluorescence polarization (Relative Polarization_{corr}) in CAF-CM/C16:0-treated HCT116 cells. **B**, Wound healing assay of HCT116 cells incubated with CAF-CM and treated with the indicated compounds as detected by IncuCyte ZOOM. n=3. **C**, Quantification of the wounding rates of HCT116 cells in **(B)**. # and \$ indicate comparisons with the corresponding

control groups. Wound width was normalized to the initial width when the wound was created. n=3. **D**, The number of crystal violet-stained cells among HCT116 cells that invaded the Transwell. n=3. Bars, mean \pm SD. n=3 *,\$ p < 0.05; **, ##, \$\$ p < 0.01; ***, ###, \$\$\$ p < 0.001.

Figure S4

DLD1



Figure S4. S-FAs inhibit CRC cell migration. Wound healing assay of DLD1/HCT116 cells incubated with CAF-CM and treated with the indicated compounds as detected by IncuCyte ZOOM. n = 2 independent experiments



with 2 CAF-CM samples from different CAF cell lines.



CM or control medium, as visualized by 2-NBDG fluorescence. n=3. **B**, Relative levels (% of control) of glucose uptake and lactate production in HCT116 cells after incubation with CAF-CM. n=3. **C**, Impact of CAF-CM incubation on the extracellular acidification rate (ECAR). The ECAR of HCT116 cells was monitored using a Seahorse XF24 analyzer. n=2 independent experiments with similar results. **D-E**, Glucose uptake (D) and lactate production (E) in CAF-CM-treated HCT116 cells cultured in the absence or presence of C16:0 (concentration from 0 to 50 μ M). n=3. **F**, The protein level of GLUT4 in CAF-CM-treated DLD1 cells was assessed by Western blotting. n=3. **G**, The viability of HCT116 cells incubated with CAF-CM was assessed by annexin-V/PI assay after 48 h incubation with 3-BrPA. The value in each panel indicates the % of surviving cells. n=2 independent experiments with similar results. Bars, mean ± SD. *, # p < 0.05; **, ##, \$\$ p < 0.01; ***, \$\$\$ p < 0.001.





Figure S6. CAF-derived lipids boosted CRC growth and metastasis.

A, The proliferation rate of HCT116^{low-SCD} cells, as evaluated by phase object confluence (%) with IncuCyte ZOOM. n=3. **B**, HCT116 cells treated as indicated were incubated with BODIPY 500 (lipid uptake). The cells were analyzed by flow cytometry. **C**, Western blot showing CD36 expression in HCT116 cells incubated with CAF-CM. **D**, The HCT116 cells treated with indicated CM were incubated with BODIPY 500/510 to analyze lipid uptake by flow cytometry. Bars, mean \pm SD. ***p < 0.001; NS, not statistically significant.



Figure S7. Treatment of cells with C16:0 increased lipotoxicity-induced CRC cell apoptosis. A, Growth curve of HCT116 cells treated with the indicated compounds. IncuCyte ZOOM software was used to process the image data for phase object confluence. n=3. **B**, C16:0 induces apoptotic cell

death in HCT116 cells. Cell viability assay of HCT116 cells treated with different concentrations of C16:0 (0, 50 and 100 μ M) for 48 h. Apoptotic cells were identified by propidium iodide (PI) and annexin V staining. n=3. **C**, Reactive oxygen species (ROS) levels in HCT116 cells with or without CAF-CM incubation and SCD inhibitor treatment were detected by flow cytometry. n = 2 independent experiments with similar results. **D**, SCD inhibitor treatment increased HCT116 cells sensitivity to C16:0. Cell viability was assessed by annexin-V/PI assay. n=3. **E**, Relative protein level of BIP (GRP78 BiP) in HCT116 cells incubated with CAF-CM or after C16:0 treatment (50 μ M, ~6 h) determined by Western blotting. Anti-BIP antibody (1:1000 dilution; Abcam, Cambridge, MA, USA), n=3. **F**, HCT116 cells incubated with CAF-CM and then treated with C16:0 were detected by oil red/hematoxylin staining. The cells were imaged using an inverted microscope. n = 3. **G-H**, FACS was used to detect the apoptosis of the indicated cells. Bars, mean ± SD. *, p < 0.05; **, p < 0.01; ***, p < 0.001.



Figure S8



different concentrations of C16:0 (0, 50, 100 and 200 μ M) for 48 h and then detected by the annexin-V/PI assay. Apoptotic cells were identified by propidium iodide (PI) and annexin V staining. n=3

Figure S9







Figure S9. C16:0 acts synergistically with 5-FU to effectively kill CRC cells incubated with CAF-CM *in vitro*. A, FACS was used to detect the apoptosis of the indicated cells. DLD1 cells were treated with the indicated

concentration of C16:0/5-FU for 48 h and subjected to the annexin-V/PI assay. **B**, C16:0/5-FU induced apoptotic cell death in HCT116 cells. n=3. **C**, FACS was used to detect the apoptosis of the indicated cells. HCT116 cells were treated with the indicated concentration of C16:0/5-FU for 48 h and subjected to the annexin-V/PI assay. Bars, mean \pm SD. *, p < 0.05; **, p < 0.01; ***, p < 0.001.