Figure S1. The expression of m6A related genes in CESC and HNSC and their
 location.

3 (A) Venn of differentially expressed m6A regulators in tumor tissue and normal tissue

4 in CESC and HNSC. (B) Expression levels of the 17 shared differentially expressed

5 m6A regulators. (C-D) The location of DEGs in HNSC and CESC on chromosomes.

6 Figure S2. Expression levels of m6A regulators in HPV-positive and negative7 HNSC.

8 Figure S3. Expression spectrum of m6A regulators in HPV-positive and negative
9 CESC and survival analysis in HPV-related cancer.

(A) Expression profiles of m6A regulators in HPV-positive and negative CESC. (B)
Venn diagram of m6A regulators differentially expressed in tumor tissue and normal
tissue in CESC and HNSC and HPV-positive and negative HNSC. (C) DFS survival
map of m6A regulators. (D) Kaplan-Meier curve of m6A regulators in HNSC. (E)
Kaplan-Meier curve of m6A regulators in CESC.

15 Figure S4. Overall survival analysis of m6A regulators in HPV-related cancer.

16 (A)Overall survival map of m6A regulators. (B) Kaplan-Meier curve of m6A

17 regulators in HNSC. (C) Kaplan-Meier curve of m6A regulators in CESC.

18 Figure S5. The landscape of genetic variations of m6A regulators in HNSC and19 CESC.

(A) Overview of the types of mutation observed. (B) Breakdown of the observed
substitution mutations. (C) Alteration frequency in HNSC and CESC. (D) Patients
were divided into two groups with or without alterations on the m6A regulators. The

- 23 graph shows genes with the highest frequency in any group. (E) Mutation count,
- 25 Figure S6. METTL3 was involved in tumor immune response in HPV-related

fraction genome altered, and MSIsensor scores in any group.

26 cancer.

24

- Patients were ranked according to METTL3 expression, and the top 50% of patients
 were in METTL3 ^{high} status, otherwise, they were in METTL3^{low} status. Enrichment
 analysis and visualization of the filtered DEGs were performed with clusterProfiler,
 org.Hs.eg.db, and the ggplot2 R package (R 3.6.3). (A-B) Enrichment analysis of
 differential genes in CESC. (C-D) GSEA analysis of differential genes in immune
 response-related pathways. (E-F) Enrichment analysis of differential genes in HNSC.
- 33 Figure S7. Correlation of immune checkpoint molecules with METTL3
 34 expression in HPV-related cancer.
- 35 (A) Relative mRNA expression level in C33A. (B) Correlation of immune checkpoint
 36 molecules and METTL3 expression in HNSC. P values are indicated as *, P < 0.05;
- 37 **, P < 0.01; ***, P < 0.001; and ****, P < 0.0001.
- **38 Figure S8. Correlation of immune cell infiltration with METTL3 expression in**
- **39 HPV-associated cancers.**
- 40 (A) Correlation analysis of the number of immune cells and METTL3 expression in
- 41 CESC using TIMER 2.0. (B) Correlation analysis of the number of immune cells and
- 42 METTL3 expression in HNSC using TIMER 2.0.
- 43 Figure S9. Immune cell infiltration levels in HPV-related cancer.
- 44 Patients were ranked according to METTL3 expression, and the top 50% of patients

45	were in METTL3 ^{high} status, otherwise, they were in METTL3 ^{low} status. (A) The level
46	of immune cell infiltration in CESC. (B) The level of immune cell infiltration in
47	HNSC. P values are indicated as *, P < 0.05; **, P < 0.01; ***, P < 0.001; and ****, P
48	< 0.0001.
49	Figure S10. Clinical and immune relevance of METTL3 to OSCC.
50	(A-B) The relationship between METTL3 and the prognosis of OSCC. (C) The
51	diagnostic predictive value of METTL3 in OSCC. (D) Correlation between METTL3
52	expression and immune cells infiltration. (E-F) Correlation between the expression of
53	immune checkpoint molecules and the expression of METTL3.
54 55 56	Supplementary Table 1: Sequence of primers used in real-time PCR.
57	







Over-expressed Genes
 Under-expressed Genes
 The gene positions are based on GRCh38.p2(NCBI). 12496 Genes.















Α









Primer	Sequence (5'to3')
TNFSF9-F	GGCTGGAGTCTACTATGTCTTCT
TNFSF9-R	ACCTCGGTGAAGGGAGTCC
BTN2A2-F	GGGCCAGCTAATCCCATCC
BTN2A2-R	GGTGATTCTTCCCCGGTACTC
CD274-F	GCTGCACTAATTGTCTATTGGGA
CD274-R	AATTCGCTTGTAGTCGGCACC
CD47-F	TCCGGTGGTATGGATGAGAAA
CD47-R	ACCAAGGCCAGTAGCATTCTT
PVR-F	GGACGGCAAGAATGTGACCT
PVR-R	GGTCGTGCTCCAATTATAGCCT
METTL3-F	AGATGGGGTAGAAAGCCTCCT
METTL3-R	TGGTCAGCATAGGTTACAAGAGT
GAPDH-F	ACAACTTTGGTATCGTGGAAGG
GAPDH-R	GCCATCACGCCACAGTTTC
KIR2DL4-F	TCCCTGTCCCTGAGCTCTAC
KIR2DL4-R	AAGGTCACGTTCTCTCCTGC
CD86-F	AGGCAACAATGAGCAGACCA
CD86-R	ACTATGGCTTGTTGGGTGGG
VTCN1-F	CTGCCTCTCAGCCCTTACCT
VTCN1-R	GCCTTCTGCTTTTGGCTTCTT

Supplementary Table 1: Sequence of primers used in real-time PCR.